

Primerdesign™ Ltd

Vesicular stomatitis virus

genesig® Easy Kit

for use on the genesig® q16

50 reactions


GENESIG

Kits by Primerdesign


For general laboratory and research use only

genesig[®] Easy: at a glance guide


For each RNA test

Component	Volume	Lab-in-a-box pipette	
VSV primer/probe mix	5 µl	●	
Your RNA sample	5 µl	●	
oasig OneStep Master Mix	10 µl	●	

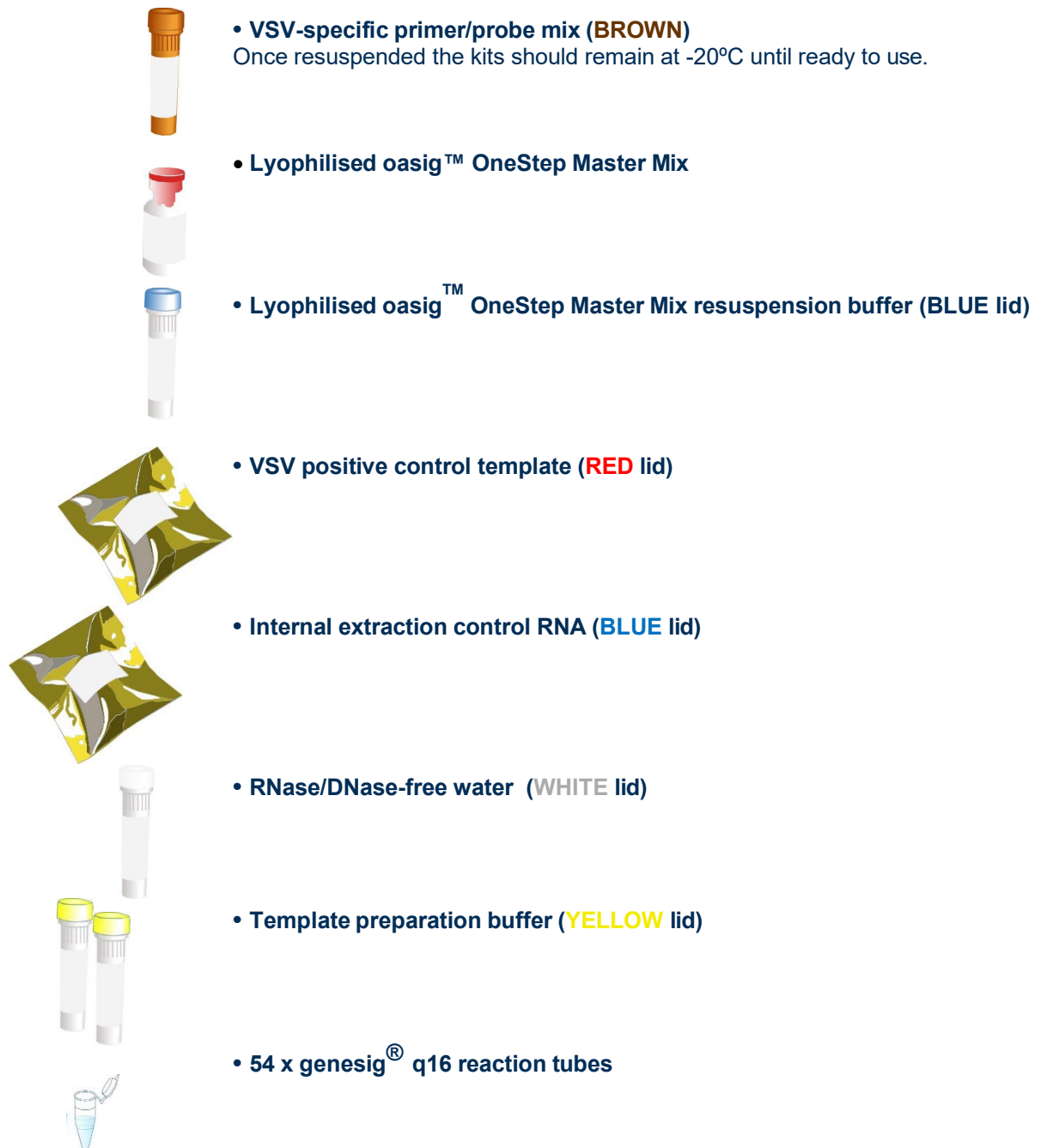
For each positive control

Component	Volume	Lab-in-a-box pipette	
VSV primer/probe mix	5 µl	●	
<u>Positive control template</u>	5 µl	●	
oasig OneStep Master Mix	10 µl	●	

For each negative control

Component	Volume	Lab-in-a-box pipette	
VSV primer/probe mix	5 µl	●	
<u>Water</u>	5 µl	●	
oasig OneStep Master Mix	10 µl	●	

Kit Contents



Reagents and equipment to be supplied by the user

genesig® q16 instrument

genesig® Easy Extraction Kit

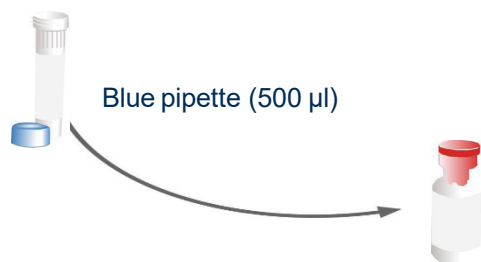
This kit is designed to work well with all processes that yield high-quality RNA and DNA, but the genesig Easy extraction method is recommended for ease of use.

genesig® Lab-In-A-Box

The genesig Lab-In-A-Box contains all of the pipettes, tips and racks that you will need to use a genesig Easy kit. Alternatively, if you already have these components and equipment, these can be used instead.

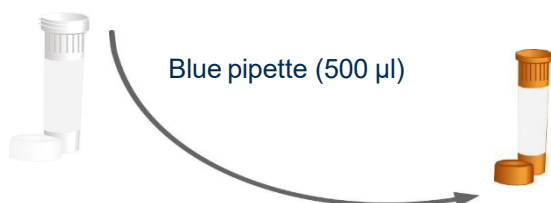
Step-by-step guide

1. Resuspend the test components



Use the blue pipette to transfer 500 µl* of the oasis OneStep Master Mix resuspension buffer into the tube of lyophilised oasis OneStep Master Mix and mix well by gently swirling.

*Transferring 525 µl of the oasis OneStep Master Mix resuspension buffer to your oasis OneStep Master Mix (instead of the 500 µl recommended above) will enable you to take full advantage of the 50 reactions by accounting for volume losses during pipetting. In order to do so with the genesig Easy fixed volume pipettes use 1x blue, 2x red and 1x grey pipettes to make the total volume. Please be assured that this will not adversely affect the efficiency of the test.



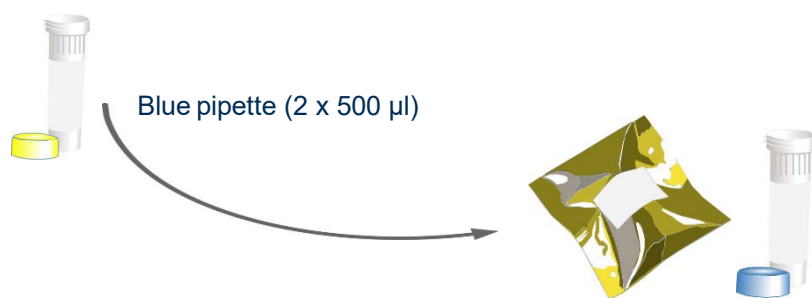
Then use the blue pipette to transfer 500 µl of water into the brown tube labelled VSV primers/probe. Cap and shake tube to mix. A thorough shake is essential to ensure that all components are resuspended. **Failure to mix well can produce poor kit performance.**

These components are now ready to use. Store them in the freezer from hereon.

Top tip

- Ensure that the primer/probe mix is mixed thoroughly before each use by shaking.
- Once resuspended do not expose genesig Easy kit to temperatures above -20°C for longer than 30 minutes at a time.

2. Internal extraction control



Use the blue pipette to transfer 1000 µl (2 x 500 µl) of template preparation buffer into the Internal Extraction Control RNA tube. Cap and shake tube to mix.

Your kit contains Internal Extraction Control RNA. This is added to your biological sample at the beginning of the RNA extraction process. It is extracted along with the RNA from your target of interest. The q16 will detect the presence of this Internal Extraction Control RNA at the same time as your target. This is the ideal way to show that your RNA extraction process has been successful.

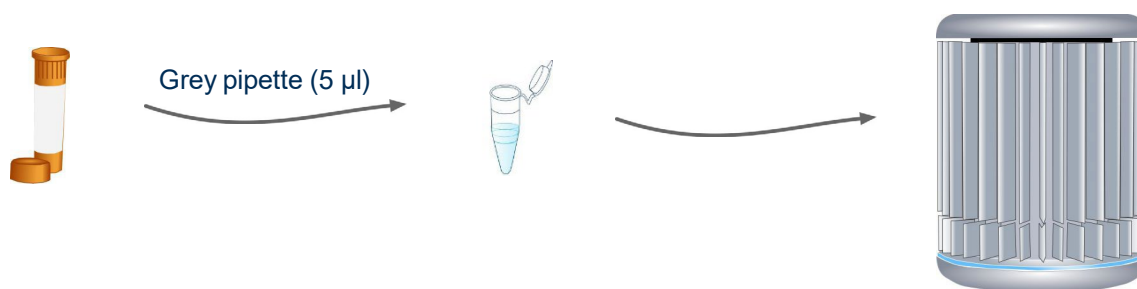
If you are using an alternative extraction kit:

Use the red pipette to transfer 10 µl of Internal Extraction Control RNA to your sample **after** the lysis buffer has been added then follow the rest of the extraction protocol.

If using samples that have already been extracted:

Use the grey pipette to transfer 5 µl of Internal Extraction Control RNA to your extracted sample.

3. Add primer/probe mix to all reaction tubes

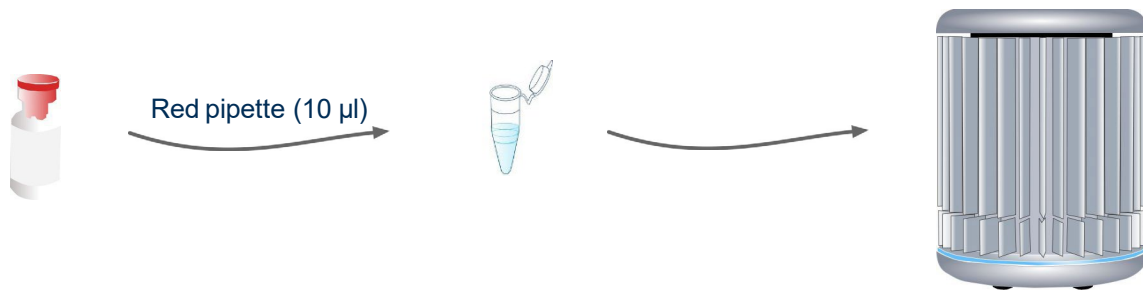


For every reaction to be run, use the grey pipette to add 5 µl of your VSV primers/probe mix to every tube.

Top tip

- Always pipette the primer/probe mix directly into the bottom of the tube.
- You can label the tube lids to aid your reaction set-up but avoid labelling tube sides.

4. Add Master Mix to all reaction tubes



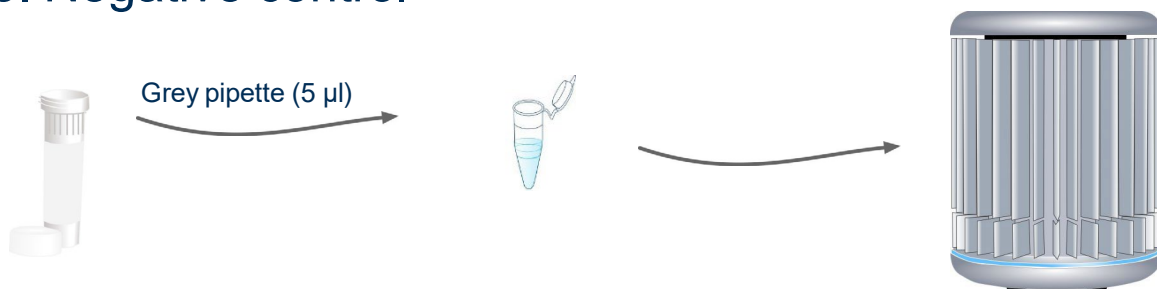
For every reaction to be run, use the red pipette to add 10 µl of the oasis OneStep Master Mix to the tubes containing primer/probe mix.

Move swiftly to begin your q16 run, as any delay after the oasis OneStep Master Mix has been added can affect the sensitivity of your test.

Top tip

- Always add the oasis OneStep Master Mix to the side of the tube to reduce the introduction of bubbles.

5. Negative control



For each test, you will require a negative control. Instead of RNA, water is used. This sample should typically prove negative, thus proving that all of your positive samples really are positive.

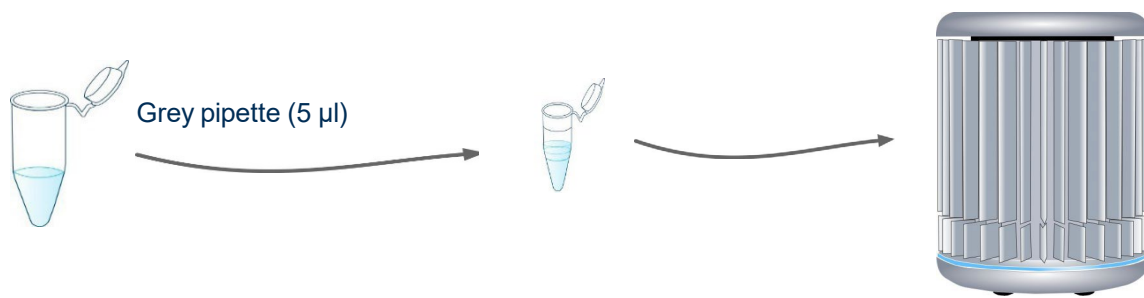
To create a negative control reaction, simply use the grey pipette to add 5µl of the water to the required reaction tubes. Close these tubes after adding the water.

Because some genesig kit targets are common in the environment, you may occasionally see a “late” signal in the negative control. The q16 software will take this into account accordingly.

Top tip

- Always add the water to the side of the tube to reduce the introduction of bubbles.

6. Set up a test

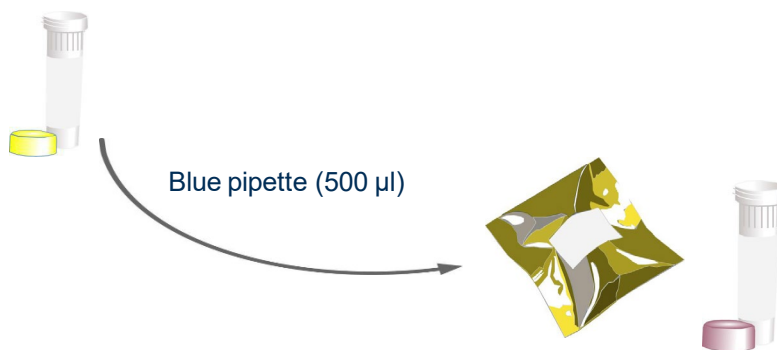


For each sample you wish to analyse, use the grey pipette to add 5 µl of your RNA sample to the required reaction tubes. Close these tubes after adding the sample. Always change pipette tips between samples.

Top tip

- Always add the RNA sample to the side of the tube to reduce the introduction of bubbles.

7. Positive control

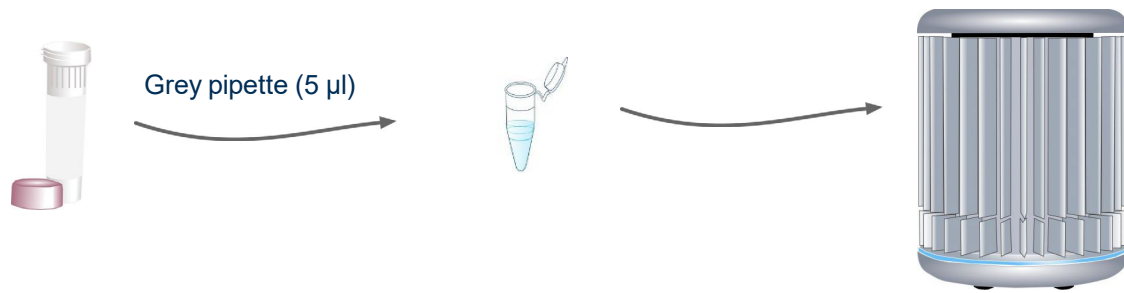


Use the blue pipette to transfer 500 µl of template preparation buffer into the positive control template tube. Cap and shake tube to mix.

Each time you run a test, you will require a positive control. This is a small portion of RNA from your target of interest. It serves two purposes:

1. It will always test positive so it shows that everything is working as it should be.
2. The q16 software knows how much RNA is present in the positive control. So, it can automatically compare your sample of interest with the positive control to calculate the amount of target RNA in your sample.

To create a positive control reaction, simply use 5 µl of the positive control instead of your RNA sample.



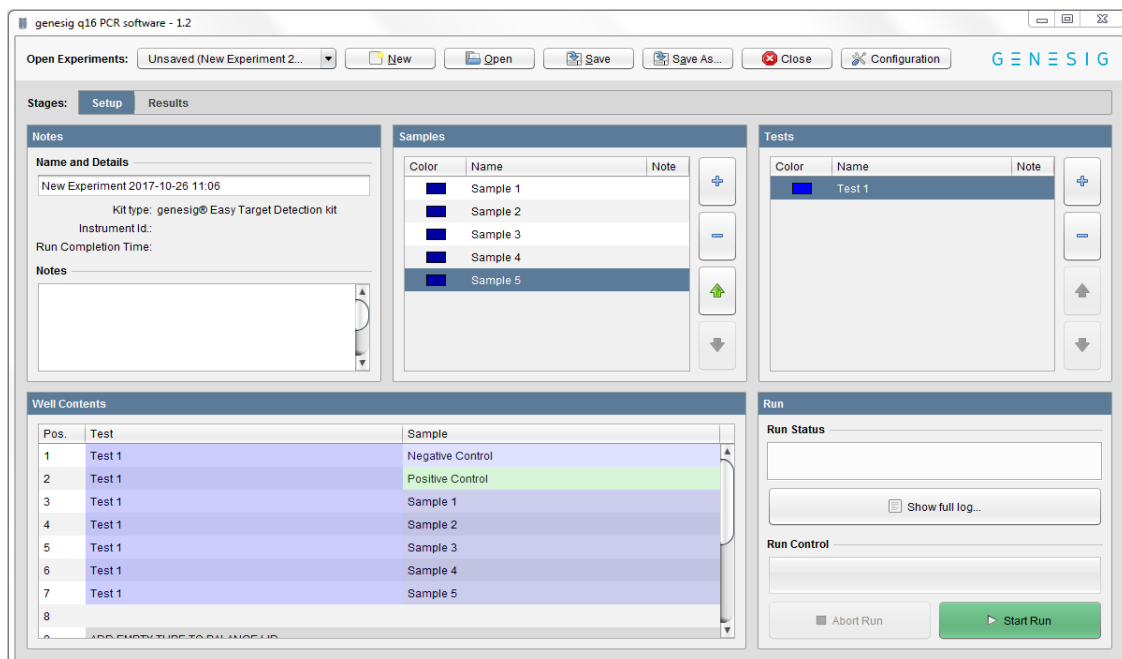
Take great care when setting up your positive control. The positive control template has the potential to give you a false positive signal in your other samples. Set up positive controls last after all other sample tubes are closed. Always change pipette tips between samples. You may even choose to set up positive controls in a separate room.

Top tip

- Always add the positive control template to the side of the tube to reduce the introduction of bubbles.

8. Running the test

Place the tubes into the correct positions in your q16 as defined by the software; this may include positioning empty tubes to ensure that the q16 lid is balanced. The run can then be started.



Top tip

- Before loading tubes into the q16, check for bubbles! Flick the bottom of the tubes to remove any bubbles that may have formed during the test set-up.
- Apply centrifugal force with a sharp wrist action to ensure all solution is at the bottom of the reaction tube.
- When repeating a test, you can use a previous file as a template by clicking 'open' then selecting File name > Files of Type > Experiment file as template

What do my results mean?

Analysis of your data is carried out automatically by the genesig q16. The following information is designed to help you fully understand a result or to troubleshoot:

“Positive”

Explanation

Your sample has produced a positive result. Your target of interest is present, and you can use the reported quantity.

“Negative”

Explanation

Your sample has produced a negative result. The target is not present in your sample.

“Test contaminated”

Explanation

The Negative Control should be completely free of any DNA/RNA. If you see this error message, it means that at some point during the set-up, the Negative Control has been contaminated with DNA/RNA and has given a positive signal. This contamination has invalidated the test. The Positive Control and your test samples are both possible sources of contaminating DNA/RNA. The genesig q16 reaction tubes from previous runs will also contain very high amounts of DNA, so it is important that these are carefully disposed of after the run is completed and NEVER OPENED. It may be the case that your kits have become contaminated, which will lead to the same problem occurring repeatedly.

Solutions

1. Clean your working area using a commercial DNA remover solution to ensure the area is DNA free at the start of your run and re-run the test.
2. If the problem persists, then the kit has become contaminated, and it will have to be discarded and replaced with a new kit. When you open the new kit, run a simple test to show that changing the kit has solved the problem. Prepare a test, which includes only the Positive Control, the Negative Control and one 'mock sample'. For the 'mock sample' add water instead of any sample RNA. The result for the Negative Control and the mock sample should be negative, indicating that contamination is no longer present.

Preventive action

An ideal lab set-up has a 'Clean area' where the test reagents are prepared and a 'sample area' where DNA/RNA samples and the Positive Control template are handled. The best workflow involves setting up all the test components (excluding the positive control template) in the clean area and then moving the tests to the sample area for the sample and Positive Control addition.

“Sample preparation failed”

Explanation

The test has failed because the quality of the sample was not high enough. The Internal Extraction Control component identifies whether the sample has been prepared correctly and is of suitable quality. This error message means that this quality control test has failed, and the sample quality is not high enough for analysis.

Solutions

1. Check the sample preparation protocol for any user errors, then repeat.
2. Quality samples can result from overloading the sample preparation protocol with too much starting material. Try reducing the amount of starting material, then repeat.
3. Failing to add the Internal Extraction Control RNA to your sample during the sample preparation protocol can also lead to a reported result of “sample preparation failed”. Ensure that this step has not been overlooked or forgotten. If your samples are derived from an archive store or from a process separate from your genesig Easy extraction kit; you must add 5 µl of Internal Extraction Control RNA into each 0.5 ml of your sample to make it suitable for use on the q16.

“Positive result, poor quality sample”

Explanation

The test is positive, so if you are only interested in obtaining a ‘present or absent’ answer for your sample, then your result is reliable. However, the test contains an Internal Extraction Control component that identifies if the sample is of high quality. This quality control test has failed, and the sample is not, therefore, of high enough quality to accurately calculate the exact copy number of RNA present. If you require quantitative information for your sample, then proceed with the solutions below.

“Test failed”

Explanation

The test has failed because the Positive Control has not worked. The Positive Control is present to show that all aspects of the test are working correctly together. When this control test fails, the test as a whole is invalidated. This finding indicates that a problem has occurred in the reaction set-up part of the experiment and has nothing to do with sample preparation.

Solutions

1. Check the entire workflow and test set-up to look for any user errors, then repeat the test e.g., have the right colour pipettes and solutions been used with the correct tubes?
2. Ensure the positive and negative controls are inserted into the correct wells of your q16.
3. A component of the test may have ‘gone off’ due to handling errors, incorrect storage or exceeding the shelf life. When you open a new kit, run a simple test to show that changing the kit has solved the problem. Prepare a test, which includes only the Positive Control, the Negative Control and one ‘mock sample’. For the ‘mock sample’, add internal control template instead of any sample RNA. If the Positive Control works, the mock sample will now be called as a negative result.

“Test failed and is contaminated”

Explanation

The Positive Control is indicating test failure, and the Negative Control is indicating test contamination. Please read the “Test Failed” and “Test contamination” sections of this technical support handbook for a further explanation.

Solution

1. For appropriate solutions, read both the “Test failed” and “Test contaminated” sections of this handbook

Vesicular stomatitis virus

Vesicular stomatitis virus (VSV), often referred to as VSIV (for Vesicular stomatitis Indiana Virus), is a virus in the family *Rhabdoviridae*, which includes the well-known rabies virus. VSV is the prototypic member of its genus *Vesiculovirus* within the family *Rhabdoviridae*.

VSV can infect insects, cattle, horses and pigs. It has particular importance to farmers in certain regions of the world where it can infect cattle. This is because its clinical presentation is identical to the very important foot and mouth disease virus. (FMD)

The VSV genome consists of a linear, single-stranded, negative-sense RNA molecule of approximately 11 kb long, which encodes five genes: the nucleocapsid protein (N), phosphoprotein (P), matrix protein (M), glycoprotein (G) and polymerase (L). Two additional small proteins (C and C') are encoded in a second open reading frame within the P gene. Transcription of the viral genes occurs in sequential order from a single promoter at the 3' end of the genome resulting in decreasing amounts of each transcript in the order: 3' Le-N-P-M-G-L 5'

The virus is zoonotic and leads to a flu-like illness in infected humans. Natural VSIV infections encompass two steps, cytolitic infections of mammalian hosts and transmission by insects.

The main sign in animals is oral disease appearing as mucosal vesicles and ulcers in the mouth, but also on the udder and around the coronary band. Animals may show systemic signs such as anorexia, lethargy and pyrexia. The disease usually resolves within two weeks, and animals usually recover completely. Because the symptoms can also mimic those of foot and mouth disease, correct diagnosis is vital to avoid herd culling.

Serological testing is most commonly performed with an enzyme-linked immunosorbent assay (ELISA) or complement fixation, and viral isolation can also be undertaken. Nucleotide testing of the isolated virus is the most rapid and reliable method of testing.

No specific treatment is available, but some animals may require supportive fluids or antibiotics for secondary infections. Accordingly, control relies on biosecurity protocols, quarantine, isolation and disinfection to ensure the viral disease does not enter a country or herd.

Specificity

The Primerdesign genesig Kit for Vesicular stomatitis virus (VSV) genomes is designed for the *in vitro* quantification of VSV genomes. The kit is designed to have a broad detection profile. Specifically, the primers represent 100% homology, with over 95% of the NCBI database reference sequences available at the time of design.

The dynamics of genetic variation mean that new sequence information may become available after the initial design. Primerdesign periodically reviews the detection profiles of our kits and, when required, releases new versions.

The kit targets the two major sub-types of VSV, namely Vesicular stomatitis Indiana virus and Vesicular stomatitis New Jersey virus.

If you require further information or have a specific question about the detection profile of this kit then please send an email to enquiry@primerdesign.co.uk, and our bioinformatics team will answer your question.

Kit storage and stability

This lyophilised kit is stable at room temperature but should be stored at -20°C on arrival. Once the lyophilised components have been resuspended, they should not be exposed to temperatures above -20°C for longer than 30 minutes at a time and unnecessary repeated freeze/thawing should be avoided. The kit is stable for six months from the date of resuspension under these circumstances. Primerdesign does not recommend using the kit after the expiry date stated on the pack.

Suitable sample material

All kinds of sample material suited for PCR amplification can be used. Please ensure the samples are suitable in terms of purity, concentration, and RNA/DNA integrity.

Dynamic range of test

Under optimal PCR conditions, genesig VSV detection kits have very high priming efficiencies of >90% and can detect less than 100 copies of the target template.

Notices and disclaimers

This product is developed, designed and sold for research purposes only. It is not intended for human diagnostic or drug purposes or to be administered to humans unless clearly expressed for that purpose by the Food and Drug Administration in the USA or the appropriate regulatory authorities in the country of use. During the warranty period, Primerdesign genesig detection kits allow precise and reproducible data recovery combined with excellent sensitivity. For data obtained by violation to the general GLP guidelines and the manufacturer's recommendations the right to claim under guarantee is expired. PCR is a proprietary technology covered by several US and foreign patents. These patents are owned by Roche Molecular Systems Inc. and have been sub-licensed by PE Corporation in certain fields. Depending on your specific application, you may need a license from Roche or PE to practise PCR. Additional information on purchasing licenses to practise the PCR process may be obtained by contacting the Director of Licensing at Roche Molecular Systems, 1145 Atlantic Avenue, Alameda, CA 94501 or Applied Biosystems business group of the Applied Biosystems Corporation, 850 Lincoln Centre Drive, Foster City, CA 94404. In addition, the 5' nuclease assay and other homogeneous amplification methods used in connection with the PCR process may be covered by US Patents 5,210,015 and 5,487,972, owned by Roche Molecular Systems, Inc, and by US Patent 5,538,848, owned by The Perkin-Elmer Corporation.

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